

Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 15 (2005) 1005-1008

Synthesis of argentatin A derivatives as growth inhibitors of human cancer cell lines in vitro

Hortensia Parra-Delgado, Teresa Ramírez-Apan and Mariano Martínez-Vázquez*

Instituto de Química, Universidad Nacional Autónoma de México, Ciudad Universitaria, Circuito Exterior, Coyoacán C. P. 04510, México, DF

Received 4 November 2004; revised 10 December 2004; accepted 14 December 2004 Available online 13 January 2005

Abstract—The syntheses of nine argentatin A analogs are described. These compounds were assessed for their ability to inhibit growth in vitro in four human cancer cell lines. Our results showed that the presence of either a double bond at C-1/C-2, or a bromine atom or formyl moiety at C-2 as well as the presence of an isoxazol ring in argentatin A enhanced its potency in all cell lines tested. In addition, an X-ray study of (16S,17R,20S,24R)-3-oxime-20,24-epoxy-16,25-dihydroxy-cycloartan-3-one led to the determination of the correct stereochemistry of argentatin A.

© 2004 Elsevier Ltd. All rights reserved.

Triterpenoids form a large class of compounds derived biosynthetically from the cyclization of squalene with the retention of all 30 carbon atoms. Many of these naturally occurring compounds have interesting biological and pharmacological properties. For example, ursolic acid, an ursane-type triterpene, inhibits tumorigenesis, and induces tumor cell differentiation. It has also proven effective in the inhibition of angiogenesis and invasion, which are important events involved in tumor metastasis.

On the other hand, some cycloartane-type triterpenes, such as methyl quadrangularates B and D, and actein, have also shown cytotoxic activity on several cancer cell lines.⁷

Argentatin A (1), a cycloartane-type triterpene, is an important component of the resin of *Parthenium argentatum* Gray (Guayule). Guayule is a common desert shrub in Northern Mexico and Southwestern USA. It has been intensively studied as a renewable native source of natural rubber, and it is known that for each kilogram of natural rubber obtained, 1 kg of a by-product named resin is also produced. Taking into account that

the argentatins, including argentatin A, comprise 20% of the resin, it can be inferred that these compounds will be available in large amounts.

Our studies on the argentatins have demonstrated that argentatin A (1) possesses anti-microbial properties. Additionally, in a previous paper, we reported that 1 inhibited 100% of cell growth in six cancer human cell lines (HeLa, K562, U251, PC-3, MCF-7, and HCT15) at $100 \, \mu M$.

As a strategy to elucidate structural requirements involved in the cytotoxic activity of 1, we now wish to report the synthesis and biological evaluation of nine derivatives of argentatin A.

Argentatin A was obtained from the resin of *P. argent-atum* (Gray) as previously reported¹¹ and identified by comparison of its physical and spectroscopic constants (mp, ¹H and ¹³C NMR) with those reported in

Keywords: Cycloartane; Argentatin A; Cytotoxicity.

^{*}Corresponding author. Tel.: +52 55 56224403; fax: +52 55 56162203; e-mail: marvaz@servidor.unam.mx

Scheme 1. Reagents and conditions: (i) Br/CH₃COOH, 98%; (ii) (a) PhSeCl, EtOAc, (b) THF, H₂O₂ 70%; (iii) AcONa/(CH₃CO)₂O, 99%; (iv) NH₂OH·HCl, C₅H₅N, 90%; (v) CH₃COOH, CrO₃, H₂O, 40%; (vi) HCO₂Et, C₅H₅N, Na/MeOH, 89%; (vii) NH₂OH·HCl, CH₃COOH, 80%.

literature, as well as by comparison with an authentic sample.⁸

Syntheses of argentatin A derivatives (2-10) are described in Scheme 1.

The 2-bromo derivative 2 was obtained from Argentatin A (1) when treated with bromine liquid in acetic acid, while treatment of 1 with phenyl selenium chloride in ethyl acetate followed by addition of tetrahydrofuran and hydrogen peroxide yielded 3.

Compound 3 was transformed into the unsaturated acetate 4 by treatment with sodium acetate in acetic anhydride. Oxime 5 was synthesized by treatment of 1 with hydroxylamine hydrochloride in pyridine.

Lactone 6 was obtained from 1, when it was dissolved in acetic acid and treated with chromium trioxide in water.

The treatment of 1 with sodium acetate in acetic anhydride gave 7. The resulting acetate was transformed into its corresponding oxime 8 by treatment with hydroxylamine hydrochloride in pyridine.

Formyl derivative 9 was obtained from 1 by treating with sodium methoxide prepared in situ and ethyl formate. In addition, isoxazol derivative 10 was obtained from 9, when treated with hydroxylamine hydrochloride in acetic acid.

All the compounds were characterized by spectroscopic and analytical tools. Additionally, adequate crystals of 5, suitable for X-ray analysis were obtained. This analysis permitted the assignment of the structure

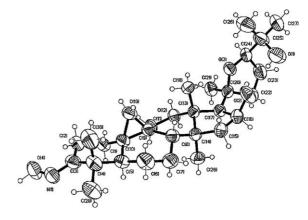


Figure 1. Crystal structure of 5.

as (16*S*, 17*R*,20*S*,24*R*)-3-oxime-20,24-epoxy-16,25-dihydroxy-cycloartan-3-one (Fig. 1).¹²

The view of molecule 5 in Figure 1 shows the *trans, cis, trans, cis* junctions for rings A–D similar configuration to those previously described for some argentatins.¹³ In 5, the ring A adopts a chair conformation, while the ring B displays a half-chair conformation due to the presence of the cyclopropane ring. The central ring, C, is present as intermediate between half chair and twist, while the five-member ring D adopts a twist conformation.

Even though argentatin A has been isolated since 1970 from *P. argentatum* and from other *Parthenium* species, ¹⁴ its stereochemistry has not been accurately determined. For instance, some authors assigned an *R* configuration to the stereogenic center at C-20, ¹⁵ while

 $IC_{50}~(SEM),~\mu M$ Compound HCT-15 (colon) K562 (leukemia) PC-3 (prostate) U251 (CNS^a) 20.22 ± 3.44 1 31.70 ± 1.10 38.61 ± 4.47 27.34 ± 1.00 2 3.23 ± 1.10 4.34 ± 0.75 11.06 ± 0.66 13.89 ± 0.44 3 16.66 ± 1.50 16.84 ± 3.46 15.26 ± 1.70 18.88 ± 0.36 >100 >100 13.93 ± 0.324 5844 + 400 35.80 ± 1.98 44.43 ± 5.57 46.91 ± 2.75 26.79 ± 2.39 6 >100 >100 >100 >100 7 40.68 ± 2.49 68.39 ± 8.30 22.69 ± 2.50 30.51 ± 3.45 8 28.87 + 3.17 15.18 ± 2.35 12.56 ± 2.42 21.37 + 1.679 9.82 ± 0.21 14.38 ± 0.78 5.69 ± 0.05 5.88 ± 1.03 10 10.24 ± 0.77 11.02 ± 1.05 13.00 ± 3.56 11.30 ± 1.55

Table 1. Effect of argentatin A derivatives on the growth of human cancer cell lines (concentration causing 50% cell growth inhibition)

Each data is given as the mean and its standard error (SEM) of at least three independent experiments. Doxorubicin: HCT-15: $0.23 \pm 0.01 \,\mu\text{M}$; K562: $0.28 \pm 0.01 \,\mu\text{M}$; PC-3: $0.32 \pm 0.02 \,\mu\text{M}$; U251: $0.09 \pm 0.02 \,\mu\text{M}$.

others did not specify the stereochemistry of the same center. Since no stereogenic center was modified in the reaction to obtain 5 from 1, then the stereochemistry of 1 was established as (16S,17R,20S,24R)-20,24-epoxy-16,25-dihydroxy-cycloartan-3-one.

The cytotoxic activity of argentatin A derivatives on K562 (leukemia), PC-3 (prostate), U251 (central nervous system), and HCT-15 (colon) cancer human cell lines was determined using the sulforhodamine B test. ¹⁶ The concentrations required to inhibit cell growth by 50% (IC₅₀) were calculated according to the protocol previously established. ¹⁶

The cytotoxic activity of argentatin A (2–10) derivatives is shown in Table 1.

An analysis of the results of cytotoxic activity of the synthetic argentatin A derivatives led to the establishment of an important fact with respect to their structure–activity relationship. The presence of either a double bond at C-1/C-2, or a bromine atom or formyl moiety at C-2, as well as the presence of an isoxazol moiety linked to ring A enhanced the potency of argentatin A in all cell lines tested (2, 3, 9, and 10 are more active than 1).

Lactone derivative **6** was inactive in all cell lines tested; this fact suggests that the presence of an isopropyl alcohol group at C-24 is essential for the inhibitory activity.

In conclusion the modification of argentatin A afforded four compounds that resulted in more active than 1 on all cell lines tested.

Acknowledgements

Partial financial support from DGAPA (IN-224802) is acknowledged. We are very grateful to Georgina Espinosa, Simón Hernández, Héctor Ríos, Nieves Zavala, Javier Pérez, Rocío Patiño, Ángeles Peña, Elizabeth Huerta, Eréndira García and Gabriela Salcedo for technical assistance. H.P.D. Acknowledges to CONACYT,

DGEP, and DGAPA-UNAM, Mexico for financial support.

References and notes

- Connolly, J. D.; Hill, R. A. Nat. Prod. Rep. 1995, 12, 609.
- 2. Tang, W.; Eisenbrand, G. Chinese Drugs of Plant Origin: Chemistry, Pharmacology, and Use in Traditional and Modern Medicine; Springer: Berlin, 1992.
- Huang, M. T.; Ho, C. T.; Wang, Z. Y.; Ferraro, T.; Lou, Y. R.; Stauber, K.; Ma, W.; Georgiadis, C.; Laskin, J. D.; Conney, A. H. Cancer Res. 1994, 51, 701.
- Lee, H. Y.; Chung, H. Y.; Kim, K. H.; Lee, J. J.; Kim, K. W. J. Cancer Res. Clin. Oncol. 1986, 120, 513.
- Sohn, K. L.; Lee, H. Y.; Chung, H. Y.; Young, H. S.; Kim, K. W. Cancer Lett. 1993, 94, 213.
- Cha, H. J.; Bae, S. K.; Lee, H. Y.; Lee, O. H.; Sato, H.; Seiki, M.; Park, B. C.; Kim, K. W. Cancer Res. 1996, 56, 2281
- 7. (a) Öksüz, S.; Shieh, H.-L.; Pezzuto, J. M.; Özhatay, N.; Cordell, G. A. Planta Med. 1993, 59, 472; (b) Smith-Kielland, I.; Dornish, J. M.; Malterud, K. E.; Hvistendahl, G.; Romming, C.; Bockman, O. C.; Kolsaker, P.; Stenstrom, Y.; Nordal, A. Planta Med. 1996, 62, 322; (c) Banskota, A. H.; Tezuka, Y.; Kim, P. H.; Qui, T. K.; Saiki, H.; Miwa, Y.; Taga, T.; Kadota, S. Bioorg. Med. Chem. Lett. 1998, 8, 3519; (d) Saxe, E. L.; Shimizu, M.; Xiao, D.; Nuntanakorn, P.; Lim, J. T. E.; Suzui, M.; Seter, C.; Pertel, T.; Kennelly, E. J.; Kronenberg, F.; Weinstein, I. B. Breast Cancer Res. Tr. 2004, 83, 221.
- (a) Céspedes, C. L.; Martínez-Vázquez, M.; Calderón, J. S.; Salazar, J. R.; Aranda, A. Z. Naturforsch. 2001, 56c, 95; (b) Komoroski, R. A.; Gregg, E. C.; Shockcor, J. P.; Geckle, J. M. Magn. Reson. Chem. 1986, 24, 534; (c) Martínez, R.; Martínez-Vázquez, M.; Zamorano-Macías, O. Org. Mass Spectrom. 1990, 25, 237.
- 9. Martínez-Vazquez, M.; Martínez, R.; Espinosa, P. G. *Fitoterapia* **1994**, *LXV*(4), 371.
- 10. Parra-Delgado, H.; Ramírez-Apan, T.; Martínez-Vázquez, M. Rev. Latinoamer. Quím. Suppl. 2001, 29, 76.
- 11. The resin, a by-product of the industrial process to obtain natural rubber from *P. argentatum* was donated by CONAZA Company (Coahuila, Mexico). The resin (157.39 g) was dissolved in a small volume of hexane and percolated over tonsil. The first fractions collected

^a Central Nervous System.

- were reunited and chromatographed on Silica gel. This procedure resulted in the isolation of argentatin A (4.2 g), which was purified by conventional procedures.
- 12. X-ray data were collected on a Siemens P4 diffractometer. Space group $P2_12_12_1$, a = 27.458 (2), b = 7.9900 (10), c = 13.0030 (10). Crystallographic data for the structure have been deposited with the Cambridge Crystallographic Data Center, CCDC-254670. Copies of the data can be obtained free of charge on application to The Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (Fax: +44 (1223)336 033; e-mail for inquiry: fileserv@ccdc.cam. ac.uk).
- 13. Martínez-Vázquez, M.; Martínez, R.; Toscano, R. A. J. Chem. Crystallogr. 1995, 25, 331.
- 14. Jakupovic, J.; Pathak, V. P.; Bohlmann, F.; Dominguez, X. A. *Phytochemistry* **1987**, *26*, 761.
- 15. Maatooq, G. T.; Hoffmann, J. H. Z. Naturforsch. 2002, 57c, 489.
- 16. (a) Monks, A.; Scudiero, D.; Skehan, P.; Shoemaker, R.; Paull, K.; Vistica, D.; Hose, C.; Langley, J.; Cronise, P.; VaigroWolff, A.; Gray-Goodrich, M.; Campbell, H.; Mayo, J.; Boyd, M. J. Natl. Cancer Inst. 1991, 83, 757; (b) Doxorubicin was used at five different concentrations (0.01–5 μM) as a positive control.